

Study of Functionalized Gold Nanoparticles with Anti-gp63 IgG Antibody for the Detection of Glycoprotein gp63 in Membrane Surface of *Leishmania* Genus Parasites

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ABSTRACT

In this work we present the preparation and functionalization of gold nanoparticles (AuNPs) for the detection of presence of gp63 glycoprotein in the surface of *Leishmania* genus parasites. AuNPs were prepared by induced laser ablation in a clean and biologically suitable media. The nanoparticles were functionalized with anti-gp63 IgG antibody in order to study the interaction with the glycoprotein component gp63 (63 kDa) present on the membrane surface of *Leishmania* genus parasites. The functionalized AuNPs showed potential as a spectrometric indicator of the parasite existence, both by the detection of the presence of the gp63 in solution and through the specific interaction with the parasites *in vitro*. The specificity of the study opens a new line of research on the use of modified nanoparticles in the development of a fast and easy assay for Leishmaniasis diagnostics.

Keywords: *Leishmania*; Nanoparticles; Surface Plasmon Resonance; gp63 Protein

1. Introduction

Nanotechnology appears to be a useful and promissory tool in biodiagnostics. The use of functionalized nanoparticles is widely applied to detect presence of biomolecules [1,2], cells and metallic ions [3-8]. Also, they have been used recently to study interactions between molecules. These devices are frequently faster, simpler and cheaper than conventional detection methods. Most of the devices developed are based in the study of Surface Plasmon Resonance (SPR) response of the Functionalized Nanoparticles (FNPs) to the changes in the local environment. This response is detected as variations in the SPR extinction band using spectrometer or in the most sensitive cases by variations in the color of the sample. The interaction of nanoparticles with organic molecules is unspecific; the specificity of the sensor is achieved by the recovering of the nanoparticle with anti-

body molecules that can interact with a very specific target. The changes observed in the extinction band of the nanoparticle recovered with the receptor molecule are due to variations in the environment surrounding the nanoparticles or the association and posterior aggregation induced by target molecules present in the media. In recent years, metallic nanoparticles are been used as vehicle to detect specific biological targets. It has been used SPR to detect the hybridization of cDNA-coated gold nanoparticles with a surface-bound RNA [9]. These cDNA-coated gold nanoparticles are used to detect ssDNA down to a concentration of 1 fM in a volume of 25 zeptomoles. Also it has been developed a method to amplify the wavelength shift observed from localized surface plasmon resonance (LSPR) bioassays in gold nanoparticle-labeled antibodies [10]. This work involves detecting surface-bound analytes using gold nanoparticle conjugated antibodies as a way to enhance SPR shifts for

more sensitive detection of low-concentration analytes. Gold nanoparticles coupled with receptor molecule (β -amiloyd) have been used as a colorimetric sensor able to demonstrate a relationship within misfolding and aggregation of proteins involved in Alzheimer's disease [2]. Gold nanoparticles coupled to lectin concanavalin A and agglutinin have also been used in order to study the specificity of this device to three cell lines expressing glycosyl complexes at their surfaces [3]. Parasite infections are spread all over the planet, especially in tropical regions, making the population under poverty or with poor conditions of life their most vulnerable target. Leishmaniosis is a vector-borne infectious disease that affects 88 countries with an estimated incidence of two million new cases each year. Leishmaniosis is caused by a protozoan parasite of the *Leishmania* genus which has the sandfly—or phlebotomous of the genus *Lutzomyia*—as its vector in the new world and Phlebotomous in the old world. About 20 *Leishmania* species are responsible for a wide range of clinical manifestations, some of them are fatal if left untreated [11]. With an expanding endemicity, an estimated 350 million people at risk and 2,357,000 disability-adjusted life years lost, leishmaniosis is becoming a worldwide re-emerging public health problem. In spite of this, its study has been neglected. One intriguing aspect of leishmaniosis is the wide spectrum of distinct clinical manifestations that include visceral, cutaneous, mucocutaneous, and diffuse cutaneous leishmaniosis [12]. Over 70 species of *Lutzomyia* are suspected to be implicated in the transmission of eight *Leishmania* species known to infect dogs in South America [13]. *Lutzomyia longipalpis* is the most common vector for visceral leishmaniosis, and as many as 100 species of the same genus are responsible for cutaneous and mucocutaneous leishmaniosis. In Venezuela, leishmaniosis control begins by having relevant epidemiological information leading to the design of a social assistance program to treat infected people. Who are they? What's their age and living conditions? And other relevant information is crucial for the success of this program. Citizens between 0 and 29 years old living in small cities or in the countryside are the most vulnerable segment of the population. In 1995 the Pan-American Health Organization reported the existence of visceral, cutaneous, and mucocutaneous leishmaniosis in Venezuela [14]. The most affected area in this country corresponds to the Andes region, where Táchira, Mérida and Trujillo states concentrate more than 60% of the infected population, with small farmers and villages population being the most vulnerable targets. Until today, the most common methods to detect Leishmaniosis infection include microscopic view, indirect immunofluorescence (using an anti-*Leishmania* IgG) confirmed by western blotting, positive response to patients after antimonial treatment (N-methylglutamine) or liposomic

B amphotericine [15], indirect immunoperoxidase method [16], polymerase chain reaction [17], serology (ELISA) and lymphoproliferative response [18]. In this way, a sensible, cheap and reliable method for the diagnosis of Leishmaniosis occurrence is of great interest in order to improve the knowledge of the illness population prevalence and help local health policy decisions. The aim of the present work is to prepare functionalized gold nanoparticles (FAuNPs) which could be able to detect a membrane surface antigen, the 63 kDa glycoprotein named as gp63 with protease activity [19,20], which is expressed on the surface of several *Leishmania* species, includes twelve different strains of *Leishmania*, including *L. major*, *L. donovani*, *L. infantum*, *L. tropica*, *L. mexicana*, *L. amazonensis*, *L. braziliensis*, and *L. enriettii* [21]. In *L. major* LEM 513, the cell is covered by approximately 5×10^5 copies of gp63 which correspond to about 1% of the total proteins of the cell [22] and the protein is anchored in the membrane by a glycopospholipid which is structurally homologous to the anchor of the variante surface glycoprotein (VSG) of *Trypanosoma brucei* [19]. Two reports that analyse the peptide maps of iodinated gp63 of *L. donovani*, *L. major*, and *L. tropica*, and *L. donovani*, *L. major* and *L. braziliensis* [21] demonstrate that the primary structures of the proteins are highly conserved. The interaction of the functionalized nanoparticles was tested in vitro. The results obtained are the first steps towards the development of a methodology able to provide very important information about Leishmaniosis occurrence in the third world countries, focusing our efforts in Venezuela.

2. Materials and Methods

2.1. Gold Nanoparticle and Antibody Binding

Mouse IgG anti-*Leishmania major* surface protease gp63 (from now on named anti-gp63) was obtained from AbD serotec OBT2004. Polyethylene glycol (PEG) from Sigma-Aldrich. Solutions were prepared using deionized water 18 m Ω filtered through 0.45 μ m Millipore filter. Gold nanoparticles were prepared by Laser Ablation of a thin film of gold immersed in an aqueous solution in PEG, using the first harmonic of a Nd:YAG laser (1064 nm) at a frequency rate of 10 Hz with a pulse duration of 8 ns and 145 mJ/pulse of power. The irradiation of the metal results in fast removal of the material that is confined to laser spot. The ejected nanoparticles remain in the liquid forming the colloidal solution that exhibited long term stability. The size-control was obtained by proper tuning of the surfactant concentration, the laser fluence and numbers of shot [23]. The size and morphology of the AuNPs were evaluated using a transmission electron microscopy (TEM) JEOL model JEM-1220 operated at 120 kV. The samples for TEM observation

were prepared by placing a drop of nanoparticle solution on colodion coated copper grids and evaporating it in air at room temperature. Images were recorded by using Kodak 4489 film. The extinction spectra of the plasmonic resonance band of the AuNPs were detected using an Ocean Optics S2000 spectrometer. The AuNPs obtained as described above are gold nanoparticles surrounded by PEG. The PEG acts like surfactant maintaining the AuNPs dispersed in the solution to avoid their aggregation; additionally the PEG permits to obtain a correct ulterior orientation of IgG. In fact, IgG will bind to the PEG by interaction at the C-terminus of the heavy chains leaving the fragment of the IgG antigen binding (FAB) towards the surrounding solution. To obtain the FAuNPs the PEG covered AuNPs were left in contact with a 1:100 anti-gp63 solution (1 mg/ml) for 24 h.

2.2. Parasite Culture

Leishmania major promastigotes were cultured at 27°C by serial passages in Schneider's insect medium pH 7.4 supplemented with 10% inactivated fetal bovine serum (FBS). Cells were harvested in late log phase ($4 - 5 \times 10^6$ parasites ml^{-1}) by centrifugation at $2500 \times g$ at 4°C for 10 minutes, and washed twice with phosphate-buffered saline (PBS, 10 mM phosphate; pH 7.4; 2.7 mM KCl; 137 mM NaCl).

2.3. Parasite Immobilization

L. major promastigotes were immobilized in a 50 ml sterile culture FALCON® flask. This was carried out by the following procedure: an L-polylysine solution (1 mg/ml) in PBS was added to the flask, enough to cover the entire flat surface, and after 30 minutes incubation, the solution discarded and the flask rinsed twice with PBS pH 7.4. A suspension of washed parasites in PBS was added to the culture flask and left for 15 minutes, or until all the parasites were adhered to the flask bottom. The cells were fixed with 3% para-formaldehyde solution in PBS for 10 minutes, washed in PBS, rinsed for 5 minutes with ammonium chloride 50 mM in PBS and finally rinsed twice with PBS.

2.4. gp63 Protein Purification

The procedure for gp63 purification was carried out as described [18] with slight modifications [21]. Briefly, 1×10^9 washed promastigotes were mixed and extracted at 0°C for 10 minutes in 10 ml of TBS containing 2% (w/v) of precondensed Triton X-114 (Sigma), and sedimented at $50.000 \times g$ for 30 minutes at 4°C. The supernatant was incubated for 5 minutes at 30°C to induce the aggregation of the Triton X-114 micelles and the solution centrifuged at $5000 \times g$ for 10 minutes at 28°C to separate the phases.

The detergent phase was diluted 1:5 with Triton X100 (Sigma) at 0.03% in Tris HCl 10 mmol/l pH 7.4, incubated for 10 minutes at 4°C and centrifuged at $5000 \times g$ for 15 minutes at the same temperature until clarification. The detergent-enriched phase containing integral membrane protein was finally diluted to 1:20 with Triton X-100 at 0.03% with Tris HCl 10 mM pH 7.4. Finally the extracted surface antigen was purified through a column of anionic exchange DEAE52 and the product analyzed by SDS PAGE to verify the purity of the sample. The purified antigen was kept in small aliquots at -20°C until used.

2.5. Light Interferometer Microscopy

Two kinds of samples were observed under the white light interferometer microscope, New View 600 model Zygo MetroPro. In one hand the control sample composed by *Leishmania* parasite cultures fixed onto glass plate which were left in contact with non-functionalized gold nanoparticles by 4 hours. In the other hand sample was composed by *Leishmania* parasite culture fixed onto glass plates and left in contact with functionalized nanoparticles by 4 hours.

3. Results

Figure 1 shows the extinction spectra of the gold nanoparticles dispersed in water with PEG as surfactant. Typical extinction band with a maximum at 524 nm is observed.

This band corresponds with the SPR of the AuNPs of approximately 30 nm in size. The inset in the figure shows the electron micrograph of the obtained particles. The size of the nanoparticles was obtained by averaging the size of the particles observed in twenty electron microscope images, assuming all to be spherical in shape.

3.1. Nanoparticles Functionalization

Figure 2 shows the corrected absorption spectra for solutions of non-functionalized gold nanoparticles (AuNPs) with PEG and the FAuNPs. This plot shows a clear red shift of approximately 4 nm in the SPR band for the FAuNPs compared with the AuNPs with PEG spectra. This red shift in the band is a typical consequence of the variation in the local environment of the nanoparticle due to the covering of its surface with anti-gp63 molecules.

3.2. Interaction of Functionalized Nanoparticles with gp63

The interaction of the AuNPs-PEG-anti-gp63 array (FAuNPs) was studied by putting in contact this array and the gp63 extract previously obtained. Solutions with

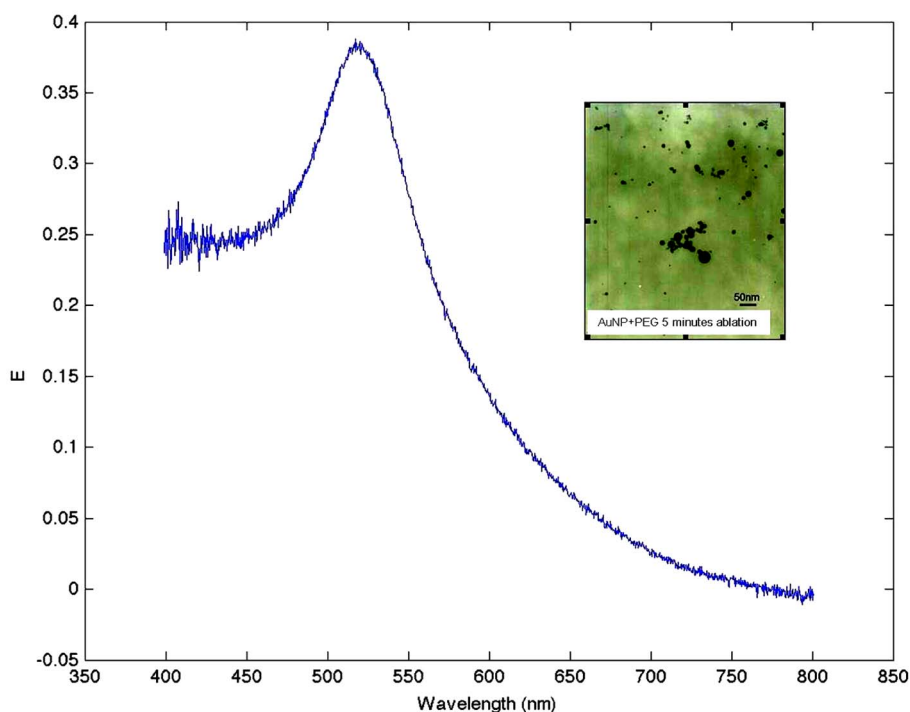


Figure 1. Extinction spectra of the gold nanoparticles dispersed in water with PEG as surfactant. In the inset electron micrograph of AuNPs.

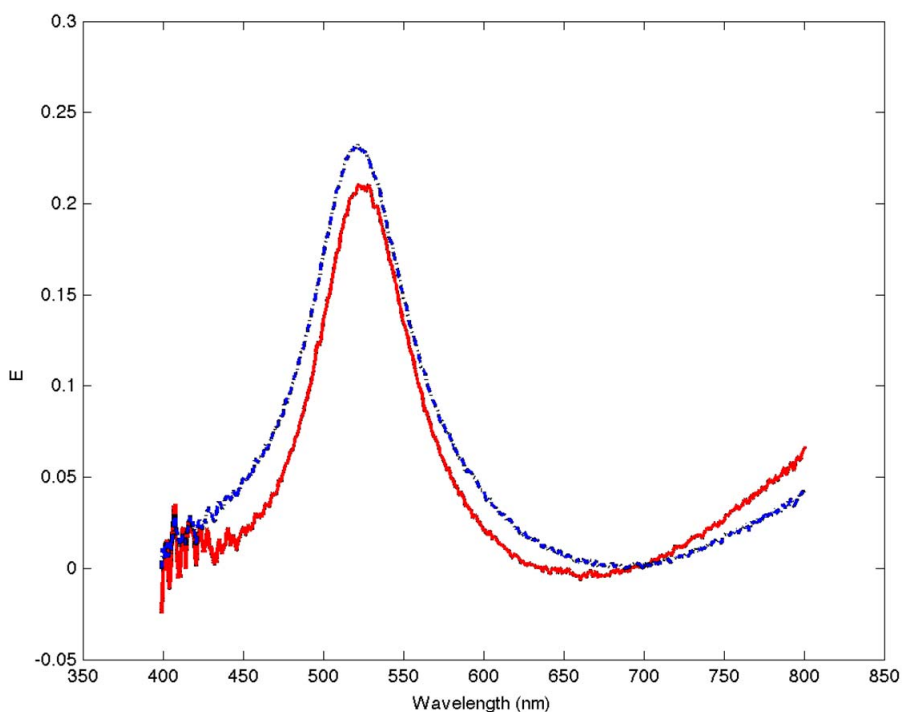


Figure 2. Corrected absorption spectra for solutions of AuNPs (blue and dashed line) with PEG and the FAuNPs (red and solid line).

5 ml of FAuNPs were mixed with additive volumes of the gp63 extract; starting from 100 μ L and increasing up to 900 μ L. A negative control represented by non-func-

tionalized AuNPs-PEG was left interact with the same gp63 antigen solution in order to discard any non specific reaction between PEG layer and the gp63 antigen. For

this negative control no variation in the extinction spectra was observed. **Figure 3** shows the corrected SPR spectra for the FAuNPs in contact with increasing amounts of gp63.

In **Figure 4** the variation of the maximum SPR intensity is plotted as a function of the quantity of gp63 added. Both plots show a decrease in the SPR signal with higher quantities of gp63 present in the media. This reduction is due to the diminishing in number of the nanoparticles responsible of this band in the media. The interaction of

the FAuNPs with the gp63 conduces to the aggregation of nanoparticles forming a new colloid of higher size and as a consequence, to the existence of fewer particles in the media that contribute to decrease in the SPR band intensity.

3.3. Interaction of Functionalized Nanoparticles with Parasites

AuNPs-PEG-anti-gp63 array was also tested *in vitro*.

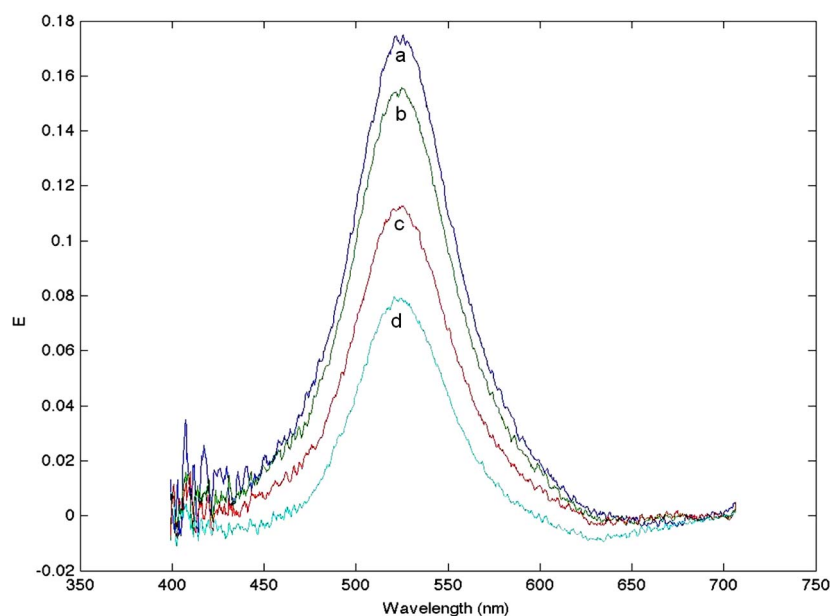


Figure 3. Corrected SPR spectra for the FAuNPs in contact with increasing amounts of gp63. Curve a: 100 μL , curve b: 200 μL , curve c: 600 μL and curve d: 900 μL .

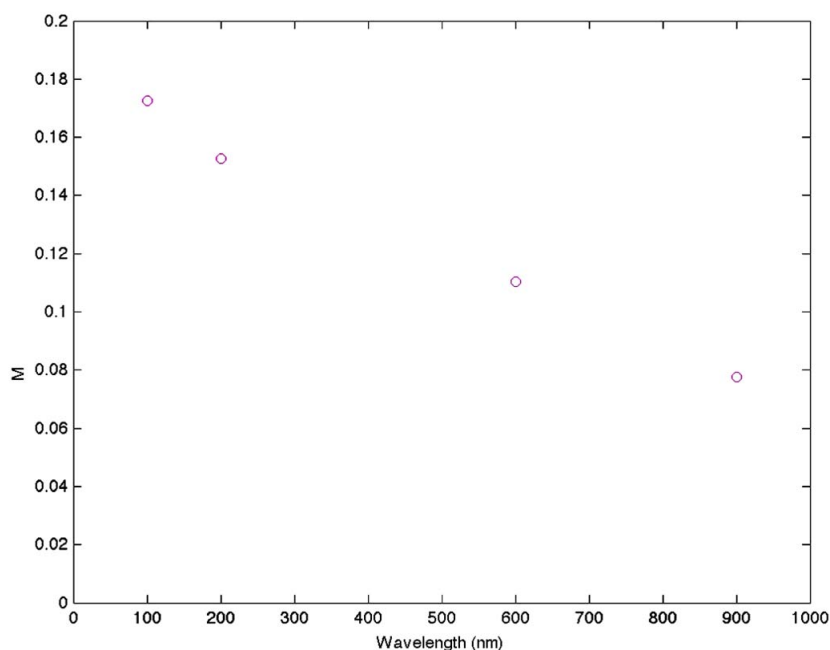


Figure 4. Variation of the maximum SPR intensity is plotted as a function of the quantity of gp63 added.

Immobilized promastigotes from *L. major* as described before were put in contact with FAuNPs and the extinction spectra were recorded every hour during 48 h at 4 °C. A conspicuous loss of the suspension color was observed. In order to determine if the observed interaction leading to observed loss of color was specific and a consequence of the antigen/antibody (gp63/anti-gp63) interaction, a blank test represented by the same arrangement without

parasite cells, was carried out obtaining no changes in the SPR spectra. Results are in **Figure 5**.

These results were confirmed using *L. mexicana* in similar conditions as before, with similar results to those obtained with *L. major*. **Figure 6** shows images of parasites supported on glass plates after exposure to non-functionalized AuNPs (AuNPs, top) and functionalized AuNPs (FAuNPs, bottom).

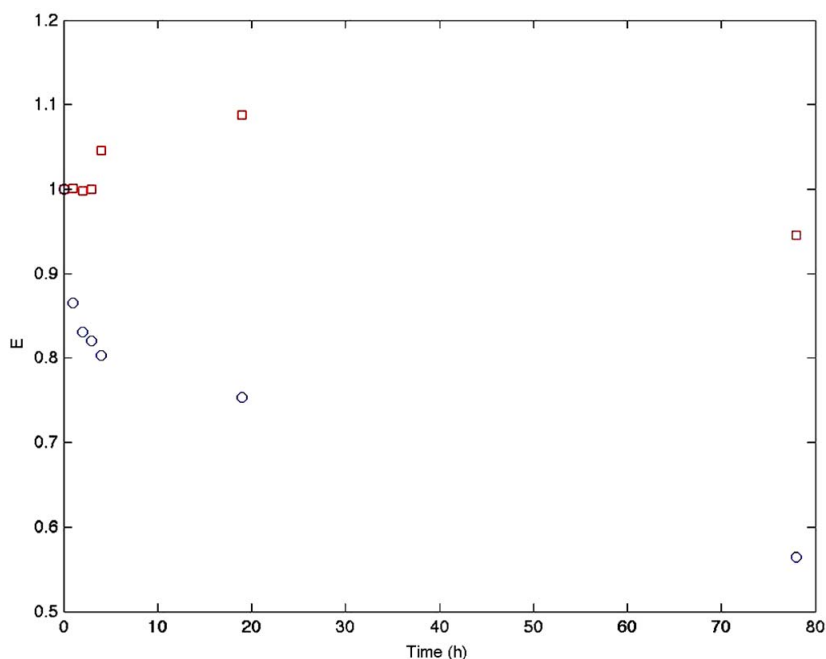


Figure 5. Variations in the SPR maxima for the FAuNPs in contact with immobilized *L. major* promastigotes cells (circles) and FAuNPs in contact with the Falcon® bottle without cells (squares).

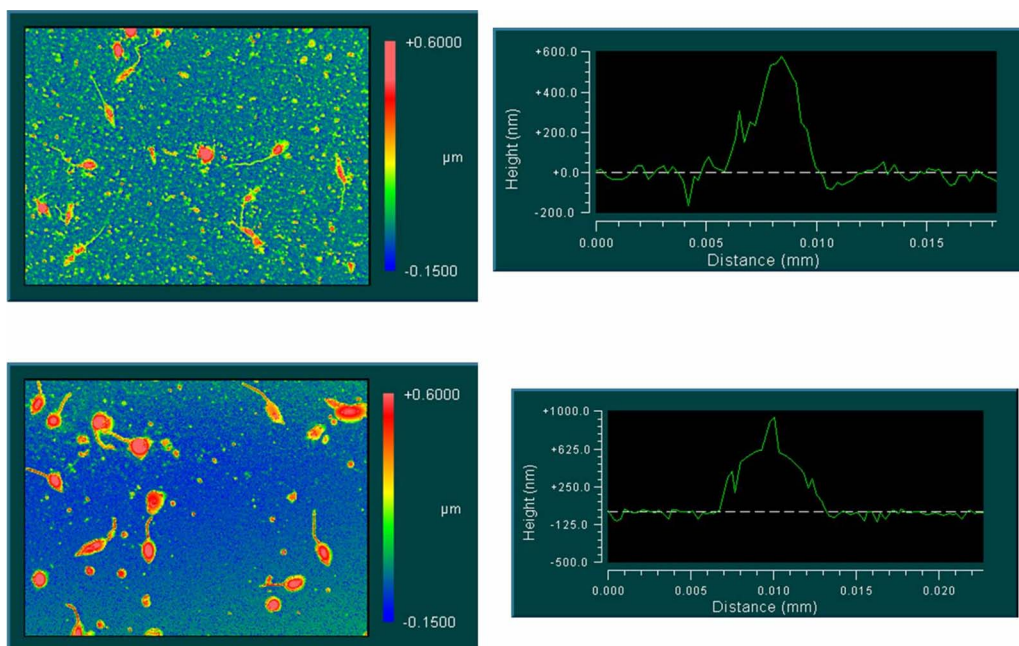


Figure 6. Interferometric images from parasites in contact with AuNPs (top) and FAuNPs (bottom).

Parasites exposed to FAuNPs are coated by metallic nanoparticles and present a greater contrast than parasites exposed to non-functionalized AuNPs. The interferometric images draw a difference in the width and height of the parasites in these two experimental conditions. The observed difference could be due to the presence of nanoparticles adsorbed on the surface of the parasite.

4. Discussion

Interaction of functionalized gold nanoparticles for the detection of *Leishmania* genus parasites was studied. The functionalized nanoparticles proved to be highly sensible and selective in the detection of the gp63 surface membrane antigen both in solution and when exposed to immobilized *Leishmania major* and *L. mexicana* promastigotes. The effect shown that can be useful as a colorimetric indicator of the parasite existence. Corrected SPR spectra for the FAuNPs in contact with increasing amounts of gp63 showed a decrease in the SPR signal with higher quantities of gp63 present in the media. The synthesis by laser ablation is a simple and easy way to use technique for obtaining metallic nanoparticles. The ablation time and the energy applied to the metallic plate are two of the parameters that must be considered when performing the synthesis. In our experiments the union between the FAuNPs to promastigote both immobilized and in suspension, as well as to protein gp63, was evidenced. These tests were designed taking advantage of the spectroscopic changes that took place due to the aggregation effect of the metallic nanoparticles [22,24,25]. The registered changes were used to expose the interaction of the FAuNPs complex with the gp63 protein. In all our experiments we observed changes in the absorption spectra as a consequence of the FAuNPs-protein interaction. In all the tests a decrease in the absorption spectrum of the AuNP occurred, in particular in the experiments using a suspension of *L. major* and a pure extract of gp63, where a displacement of the absorption peak in the normalized spectra above a wavelength of 520 nm was observed. The spectroscopic changes due to the aggregation of the metallic nanoparticles have been reported previously in the literature. The development of biosignal detection methods with the use of plasmonic nanosensors has been mentioned [26]. The authors worked with the extinction spectrum of both gold and silver nanoparticles, defining the extinction as the capacity to obscure or extinguish certain wavelength. In this sense a shift of the maxima within the extinction spectra in experiments using FAuNPs has been previously observed [22]. The arrays of immobilized proteins over metallic nanoparticles have a high potentiality for the detection of specific cells. The report of the development of applied nanotechnology for the detection of microorganisms has been mentioned

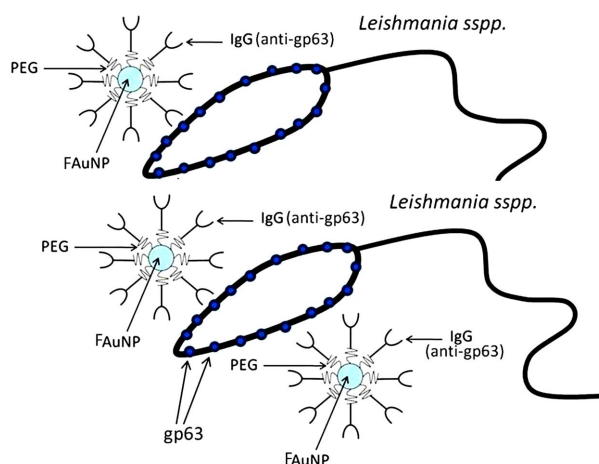


Figure 7. Model proposed for interaction between FAuNPs and *Leishmania* sp. promastigotes (PEG: Polyethylene glycol. IgG (anti-gp63): Immunoglobulin G against gp63 *Leishmania* genus protein. gp63: Glycoprotein 63 kDa from *Leishmania* genus. FAuNP: Functionalized gold nanoparticle). PS: Both promastigotes and FAuNP are not proportionally related in size.

[27], one of the methods authors mention is the spectral change due to metallic nanoparticles aggregation and that method can be used for direct detection of suspended cells, viruses or microorganism related molecules. The aggregation of the metallic nanoparticles generates a change in their geometric form. When a different tridimensional form is obtained, a change in the plasmonic resonance occurs causing a colorimetric change [24,28, 29]. This phenomenon is of interest for the present research project since our objective is to take advantage of the changes in spectral properties for the development of a biotest for the early detection of tropical parasite diseases. In the present study we have verified the capacity of the FAuNPs to couple to parasites both immobilized and in suspension. Also we have observed its ability to join two antigens in solution, as in the case of the gp63 protein present on the surface of *Leishmania major*. Taking this into account for the design of a field biotest, we would observe a discoloration of nanoparticles suspension in the presence of the parasites from samples. Taken together, our results allow us to propose the following interaction scheme between the functionalized gold nanoparticles and *Leishmania* sp. promastigotes (see **Figure 7**).

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